3

Saturday, February 22, 2014
11:22 AM

Ones that are good from PCR Check:

These are the results from February 28, 2014

Labels

- A- pNhaA+E0840 - amp
  - Promoter plus
  - ALL WRONG
- B- J23100 - amp
  - 35 bp
  - Constitutive promoter
  - WRONG
- C- K592006 - chlor
  - 250 bp
  - ALL GOOD
- D- K592016+Term - chlor
  - 1925 bp
  - D1 is good,D3 is ok
  - Digested with XP
- E- RBS - amp
  - 12 bp
  - All good
  - Digested with EX
- F- pLac + B0034 - chlor
  - 204 bp'
  - F1,F3 is good
  - Digested with SP
- G- pLac + RBS - chlor
  - This is actually the same thing as F, and G1 and G2 are good

Ran GEL:

D1, D3, E1, E2, E3, F1, F3, Ladder

- Rohan will be staying behind for the final process of cutting up the gel
- The remaining digested plasmids are kept in the fridge with the label, see Phillip for details.
- The digested plasmids may have ligated back together, use at your own discretion!

The digestion was done by Andy and Phillip

4 A DAY :)

Tuesday, March 04, 2014
3:01 PM

1. Julie

Bleached the biohazard with 10% bleach

2. Beth, Dean, Lilee

Gel extraction for D1,D3, E1, E2, E3, F1, F3

3. Greg, Edward, Chris

   Make 24 gels

- Half big wells, half small wells
- Half with seeing safe gel dye, half without it

5

Wednesday, March 05, 2014
2:26 PM

Gel extraction for D1,D3, E1, E2, E3, F1, F3

- What we have:
  - K592016+Term, RBS, pLac + B0034
- Digested:
  - C1, 2, 3
    - C1 - EX
    - C2 - EX
    - C3 - SP

WE HAVE A WORKING LABELING PROTOCOL WOOOHOOO!

-Phillip Teng

- Key:

- A- pNhaA+E0840 - amp
  - Promoter plus
  - ALL WRONG
- B- J23100 - amp
  - 35 bp
  - Constitutive promoter
  - WRONG
- C- K592006 - chlor
  - 250 bp
  - ALL GOOD
- D- K592016+Term - chlor
  - 1925 bp
  - D1 is good,D3 is ok
  - Digested with XP
- E- RBS - amp
  - 12 bp
  - All good
  - Digested with EX
- F- pLac + B0034 - chlor
  - 204 bp'
  - F1,F3 is good
  - Digested with SP
- G- pLac + RBS - chlor
  - This is actually the same thing as F, and G1 and G2 are good

10 A-Day

Monday, March 10, 2014
9:15 AM

1. Journal Notation:
Process done today:

- Gel Purification for K592006 SP and XP (useless though) they will be labeled as PDP (wrongly labeled as PP) they will be found in the fridge
- Gel purification of **J23100** (3 in 1)
    - PCR, Gel purification
    - **5 PCR tubes - B1, 2, 3, 4, 5**
    - Part names

Order of events

Things that need to be done next time:
Need to liquid purification
Need to ligate K592006
Run PCR check on B1-5
Stuff in -20 degree fridge
Derped again, sorry

Parts we need:

11

Monday, March 10, 2014
5:30 PM

Process done today:
Rachel ran gels for the PCR of J23100 and Melody did plasmid DNA extraction of the liquid culture.
Labeling: PP J23100 B1, B2, B3, B4, B5
+date

Things that need to be done next time:
J23100 needs to become a UGP at the end of tomorrow, gel ran for the 5 samples.

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results):
The gel order is the following for trial 1:

1. Ladder
2. B1
3. B2
4. B3
5. B4
6. Nothing
7. B5

The gel order is the following for trial 2:

1. Ladder
2. B1
3. B2
4. B3
5. B4
6. B5

Results:
Trial one:

Trial two:

12

Monday, March 10, 2014
5:34 PM

Process done today:
Dean and Michael: Digestion and ligation of Part A and Part B
Bethany and Leon: Transforming the DNA that was prepared previously into the cells.

Julie and Lily did the digestion for J23100 B1,B2, B3,B4, B5
Then run gels to purify these pieces

Chris (design for mascot) and Annie (design for T-shirt)
Greg prepared LB broth and autoclaved them (1000mL worth of LB with no ampicillin were made at the end of the day).
Phillip

Things that need to be done next time:
The UGP of J23100 needs to be purified.

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results)

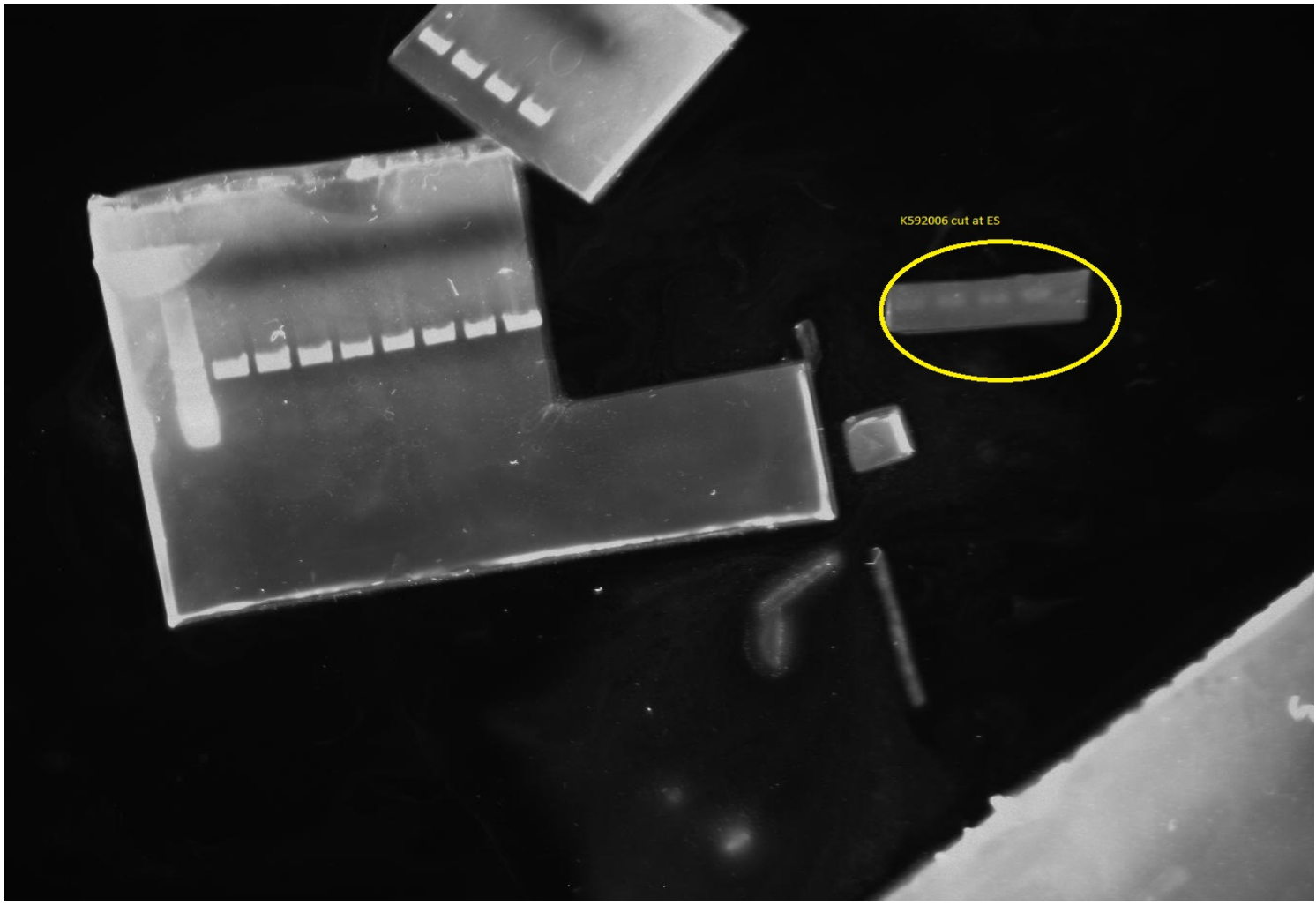
NEEDS TO BE DONE: LIGATE PART A AND B THAT WAS DIGESTED TODAY!!!!

13

Thursday, March 13, 2014
3:15 PM

Process done today:
Melody and Rachel did gel extraction for J23100 B1, B2, B3, B4, and B5.
Greg removed the LB broth from the autoclave and autoclaved some tips
Phillip ran a gel for Term A cut at XP, RBS cut at XP, and K592006 cut at ES

The gel results show that RBS and Term A failed however K592006 at ES was successful as show in the picture below:



Reflection:

The weather was very nice. But it was quite hot in the research lab. The heat was significantly bothersome enough that we felt it was more close to sweltering than pleasant.

14

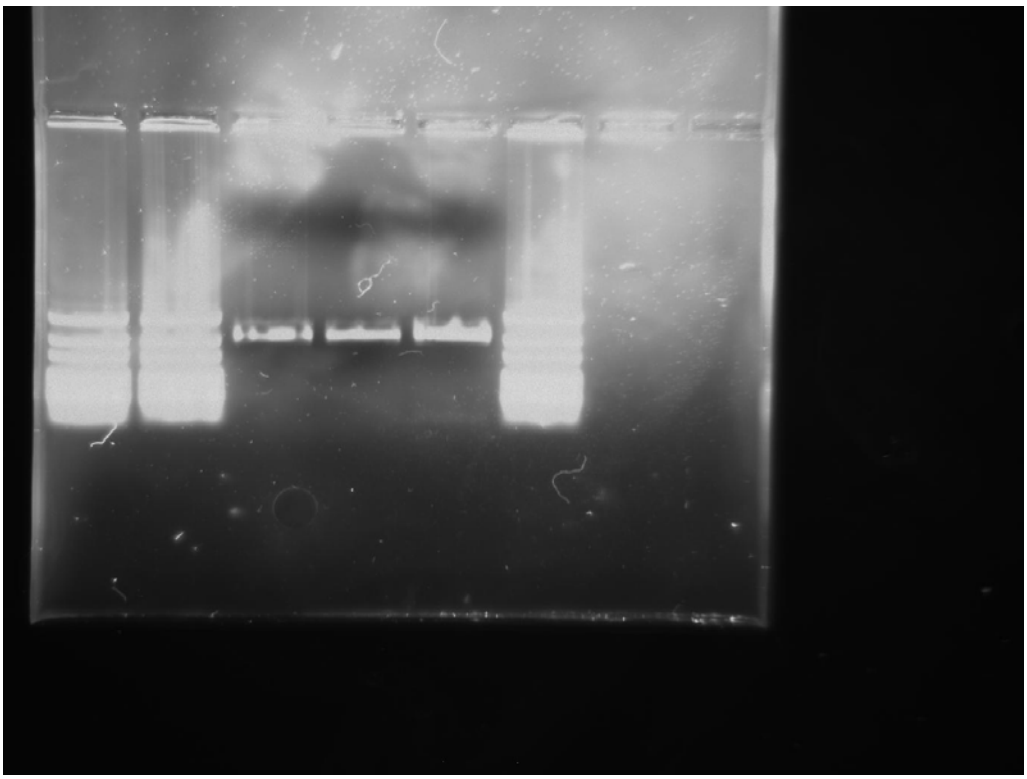
Monday, March 17, 2014
2:51 PM

Process done today:
T-shirt designs by Annie
Attempted to digest the termA and RBS at XP but the gel showed that it failed.

Things that need to be done next time:
Need to ligate K592006 with RBS at ES and EX respectively.

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results)
Failed RBS and Term A - no distinct patterns for the DNA



17

Monday, March 17, 2014
2:51 PM

Process done today:
Andy- ligation of the RBS (E1) and K592006 (EX and ES)
Rachel and Melody did transformations:
LacI, J23119, and TermA

C0012 (LacI)- found on 1M of kit plate 3
J23119 (constitutive promoter)- found on 18O of kit plate 3
B0015 (TermA)- found on 4F of kit plate
Things that need to be done next time:
3 in 1 of the process of the three parts above to see if the genes were successfully transformed.

Further plan:
Digest LacI at ES to later ligate with EX.

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results)

18 A-Day

Tuesday, March 18, 2014
3:26 PM

Process done today:
Lily, Michael- ligation of the RBS (E1) and K592006 (EX and ES)
Do not put pink eppendorf/PCR holder in water bath

The awesome transformers Leon Prime, Dean Ironhide and Bethy Bee did transformations:
LacI, J23119, and TermA

C0012 (LacI)- found on 1M of kit plate 3
J23119 (constitutive promoter)- found on 18O of kit plate 3
B0015 (TermA)- found on 4F of kit plate

Things that need to be done next time:
3 in 1 of the process of the three parts above to see if the genes were successfully transformed.

Further plan:
Digest LacI at ES to later ligate with EX.

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results)

Tuesday, May 13, 2014
2:08 PM

19

Thursday, March 20, 2014
8:46 AM

3 IN 1 For TermA, LacI, and J23119

We ran PCR for each of those and the samples were stored in the fridge.

ED Chen ligation? K592006- RBS

20

Thursday, March 20, 2014
2:44 PM

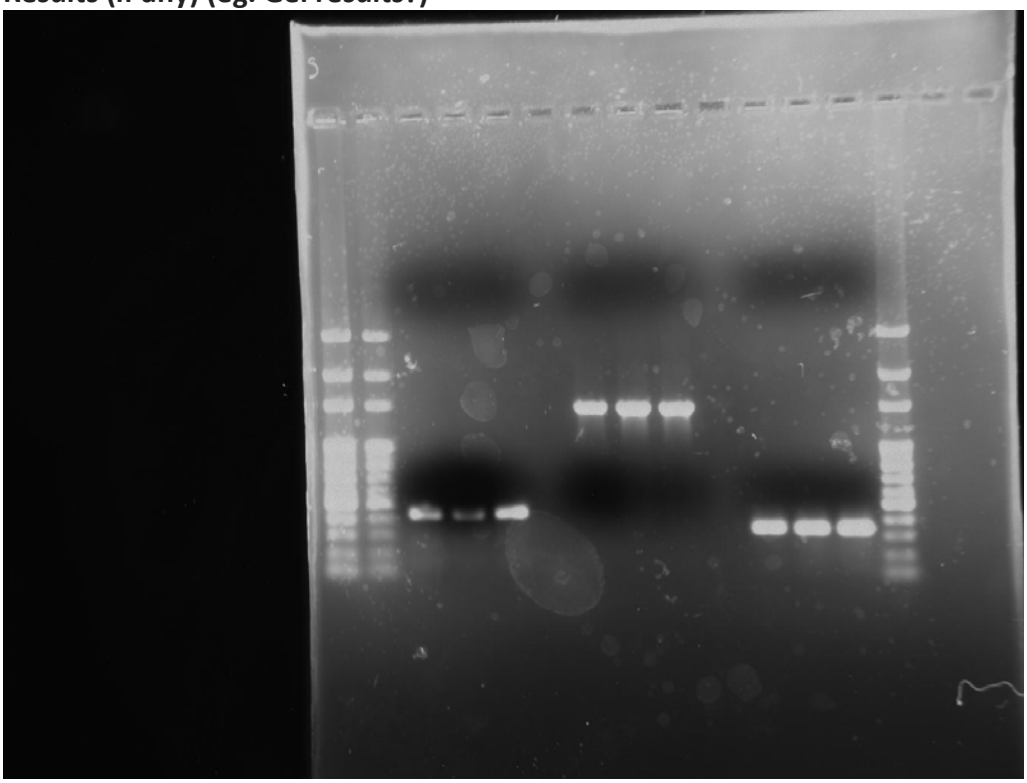
Process done today:
Phillip did 3 in 1 for RBS and K592006 (ligation seemed to be successful)
Edward, Pai, and Chris did plasmid extraction for J32119, B0015, and C0012 (9 samples)
Finished Step 54cd -

3T(Dean Ironhide and Bethy Bee) transformed the parts that need to be transformed by the transformers (CI gen, tetR gen, pCI, and pTet)

Things that need to be done next time:

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results?)



PCR check for the nine samples

J23119- 35 bp plus C= 349
B0015- 129 bp plus C= 443
C0012-

21

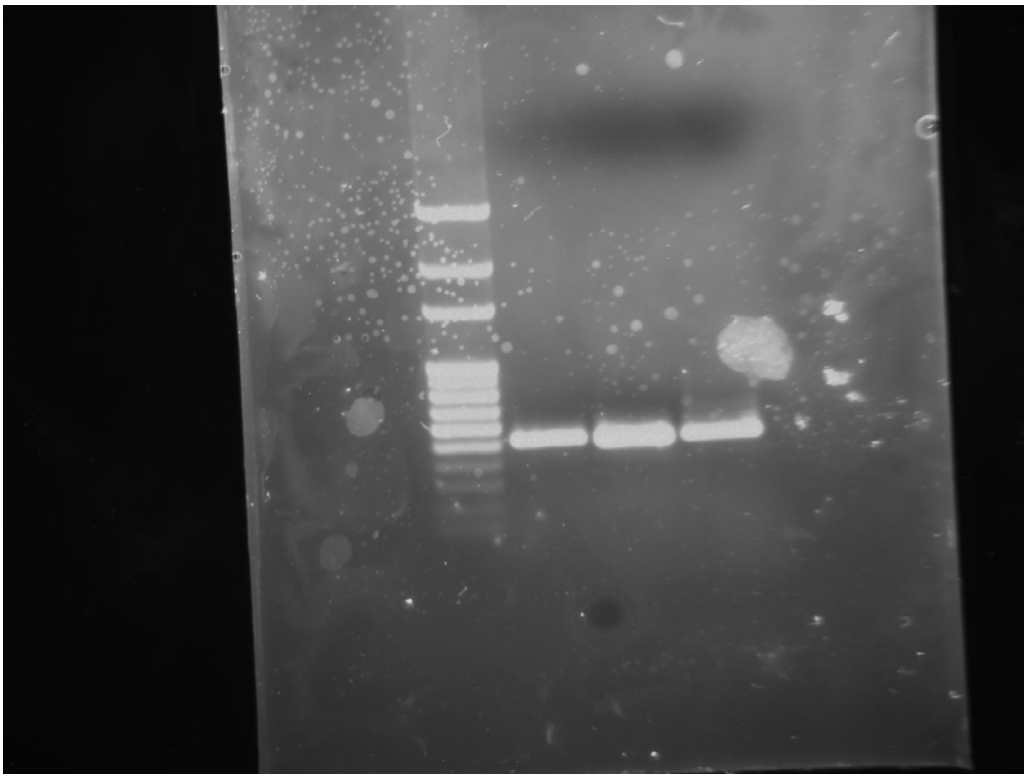
Friday, March 21, 2014
4:15 PM

Process done today:

- Digestion of J23119 at SP, C0012 at ES, b0015 at XP
- PCR check of k592006+RBS is correct
- Plasmid purified k592006+RBS
- 3-in-1 the LacI generator

- **Things that need to be done next time:**
- Continue processes

x



-

24

2014年3月22日
下午 10:09

Process that needs to be done today:

- Beth and Leon and Lily made 10 mL of amp (the most productive group of the whole class)
- Julie, Chris, Edward did plasmid extraction
    - LacI Gen P0412
    - pTet R0040
    - Gen P0451
    - TetR Gen P0440
    - PCI R0065
- Dean and Greg did 3 in 1
    - LacI Gen P0412
    - pTet R0040
    - Gen P0451
    - TetR Gen P0412
    - PCI R0065

Things that need to be done next time:

- Good question

Results (if any) (eg. Gel results? Was the transformation successful?)

*Note: A lot of the labels don't seem to match up with the actual contents. Fixed on April 7.

26

Wednesday, March 26, 2014
3:31 PM

Process done today:

- Gel purification of k592006+RBS (4 tubes)

- **Things that need to be done next time:**
- Continue processes

Format

2014年3月22日
下午 09:57

Process that needs to be done today:

Things that need to be done next time:

Notes: (eg. Where can we find the parts?)

Results (if any) (eg. Gel results? Was the transformation successful?)

Email information

Friday, March 14, 2014
5:16 PM

The research email and dropbox email is
Email: tasigem@outlook.com
Password: TASresearch

The password is the same for both he email and the dropbox account.

2014年3月22日
下午 10:11